# Chemistry Research Journal, 2018, 3(4):63-79

Available online <u>www.chemrj.org</u>



**Review Article** 

ISSN: 2455-8990 CODEN(USA): CRJHA5

# Chitwin Compounds: A New Revelation of Chemistry and Biology

# Dumitru Petru Iga

University of Bucharest, Bucharest, Rumania

**Abstract** Forms of matter showing themselves in the same relationship as objects and their mirror images were called enantiomeric, and this relationship was discovered at the level of crystals: quartz, sodium wolframate, etc. Separation of tartaric acid enantiomers, as well as the discovery of tetrahedral model of C atom, disclosed the enantiomorphous relationship at molecular level. Then the concept of chirality (kheir, hand, Gr.) partially replaced the concept of enantiomorphism. At first sight at least, the idea of chirality showed a higher mobility in physical-chemical applications. However, both points of view see their running elements – enantiomorphous or chiral forms – as separate, distinct entities. If one consider the two sides of enantiomorphism or chirality as a whole unit, one arrives at interesting and important conclusions. If a molecule contains both sides, i. e., it is formed of two enantiomeric halves (as two gloves of different handedness), the result is a symmetric or *meso* compound. Instead, if a molecule is formed of two identical sides of enantiomorphous objects (as of two gloves of the same handedness), a compound of a two-fold homochirality is obtained, since the molecules of such a compound are homochiral with one another and they are internally homochiral, at the same time. We have called the members of this subgroup *chitwin* compounds (from *chi*, chiral, and *twin*). A tentative discussion of the properties and natural distribution of *chitwin* compounds is made.

Keywords chitwin, internally homochiral, internally heterochiral, symmetry, doubling, dimerization

## Introduction

When Pasteur crystalized the salts of paratartaric acid (a racemic mixture), he noticed that he had obtained two types of crystals that were enantiomeric with one another, and separated them according to their form. (At that time the enantiomeric relationship of some crystalline compounds was relatively familiar to chemists). Then he solved them in water, and a polarimeter analysis indicated that one solution was dextrorotary and the other levorotary, with the same numerical value [1,2]. The theory of van't Hoff [3] and Le Bel [4], based on asymmetric C, as well as on spacial distribution of its bonds along the axis of a regular tetrahedron, disclosed the enantiomeric relationship at molecular level. Van't Hoff elaborated molecular models for optical isomers, however a scientific and philosophical dilemma appeared, concerning the correspondence between models and samples of different isomers [5]. This dilemma was solved with some uncertainty by E. Fischer [6-9]. Half a century after Fischer, the group of Bijvoet [10] eliminated every doubt and found out that Fischer had given the correct solution. The cornerstone of many experiments, including those of Pasteur, Fischer, Bijvoet, was tartaric acid, discovered by Scheele (1770) at the dawn of chemical sciences [11,12]. Due to these scholars, and many others, every chemistry graduate of today can state: "on this stone (tartre, Fr.) is built (stereo)chemistry". The isomers of tartaric acid were structurally correlated with the isomers of glyceraldehyde and other monosaccharides as well as with many other chiral compounds [13,14].



Kelvin [15] replaced the relationship of enantiomorphism with the relationship of chirality (kheir, hand, Gr.). It seems quite plausible that Kelvin included in this relationship other objects possessing this quality, equally familiar to the previous scholars who invented the term enantiomorphism: gloves, shoes, augers, screwdrivers, shells of some molluscs (gastropods), etc. (It's a less macabre alternative!). Kelvin also defined homochirality and heterochirality: "...two equal and similar right hands are homochirally similar. Equal and similar right and left hands are heterochirally...". Eight decades later, Prelog adopted Kelvin's chirality and other congeneric terms, however returned to the mirror and included also a reassuring mathematical term: "an object is chiral if it cannot be brought into congruence with its mirror image by translation or rotation" [16,17]. Now, armed with Kelvin's chirality and Prelog's mirror, plus a suitable amount of congruence, we may arrive at unique and important conclusions. Both Kelvin and Prelog regarded the two hands (or chiral objects) of the same handedness as separate, distinct entities. However, every couple of the same hands may be considered as a whole unit, or in chemical terms, molecules containing two (or more, but an even number of) asymmetric carbons of the same chirality, and a double set of chemical functions. Such molecules possess a twofold degree of *homochirality*: they are homochiral with one another and they are also characterized by an *internal homochirality* i. e., they are superhomochiral. Of this reason we have called them *chitwin* molecules (*chi* from *chiral*, plus *twin*).

*Chitwin* molecules constitute a distinct subgroup of the whole group of chiral ones. Their distinct character is conferred by their physical-chemical and biochemical properties. They have a unique, exceptional quality: once constructed, either of their half suffer a chemical reaction, or a series of reactions, the result is the same. Hence, they are highly versatile molecules and this versatility could have played an important role in molecular evolution, in the process of homeostasis of living things and in other fundamental natural phenomena. Every chiral compound that is not yet *chitwin* is a *chitwin* precursor. In exactly defined conditions, its doubling produces a *chitwin* derivative. As shown in this paper, *chitwin* molecules present a remarkable structural variety and they are well represented among the main classes of natural compounds: monosaccharides, amino acids, proteins, lipids, lignans and neolignans (phenolic derivatives), heterocyclic compounds, polyprenyls (isoprenoids), special natural reagents (coenzymes) and even polynucleotidic structures of palindrome type. *Chitwin* compounds constitute a real, essential, unique and trustworthy network in living matter. Moreover, *chitwin* molecules are isomers and metabolically interchangeable with two other subgroups, *meso* compounds and chiral compounds characterized by an *irregular distribution of their asymmetric centers*.

Kelvin defined heterochiral objects as a couple of chiral ones with opposed handedness, as are right hand and left hand, the two objects being seen as separate entities. If the two entities are considered as parts of the same molecular unit, *meso* substances are obtained. They are identical with one another, however they are *internally heterochiral*. Moreover, their two halves have different priorities (ranks).

In macrocosmic world, Kelvin-Prelog theory discloses us the existence of three types of objects: (i) symmetric, (ii) chiral, and (iii) chitwin ones. Every object that isn't symmetric is chiral and all we need is to confront it with Prelog's mirror and construct a replica as mirror shows it. The couple will constitute a *symmetric* system, and its two halves have different ranks (priorities). Otherwise, if we build a copy of the chiral object, their couple will constitute a *chitwin* or a homochiral system. Let's remember Pasteur's experiment: a couple of crystals of the same group constitute a homochiral (or *chitwin*) system, while a couple of enantiomeric crystals make a heterochiral (or symmetric) system. Living things also illustrate this point of view. Moreover, contrary to dead chiral objects (except hands!) that we associate just mentally – of the same, or of opposed handedness – living things of the same species are linked by the transfer of genetic material in view of species preservation – pollination by plants, and mating by animals. Symmetrical systems are illustrated by a vast number and diversity of vertebrate species. On the other hand, some species of plants (flowers) and animals (gastropods) manifest all three possibilities. A phenomenon characteristic to plants, enantiostyly, was defined as a plant sexual polymorphism in which female sex organs are deflected to the left or right, resulting in symmetrical pairs (mirror-image) of flowers [18]. It was proved that this form of sexual asymmetry has the role to promote cross-pollination in bee-pollinated plants [19,20]. The vast majority of snail species are almost exclusively of one handedness and the individuals of minority are unable to



mate with individuals of normal coil, so directionality is maintained by frequency dependent selection [21]. One can assert that these snail species are in a biological race for *chitwin* phenomenon. An exception to this rule are the snails of subgenus *Amphidromus*, because dextral (D) and sinistral (S) individuals occur in about equal proportions (so-called 'antisymmetry') in most species. It was shown that the species of this subgenus accomplish a sexual selection for dimorphism, rather than for monomorphism. Moreover, the transfer of genetic material between D and S individuals occurs more frequently than expected by chance. Physiology and biochemistry of these snails are arguments for symmetrical systems [21].

In this paper, the representation of *chitwin* molecules by the members of the main classes of natural compounds is presented in an exemplifying manner.

## 1. Monosaccharides

Monosaccharides and oligosaccharides are a good illustration of *chitwin* molecules. E. Fischer disclosed *chitwin* compounds as redox homogenous derivatives of aldoses which possessed the exceptional quality that by reversing their ends, the same product was obtained: mannose, idose, threose [6-9].

## 1.1. Redox homogenous derivatives

The group of  $C_4$  has a number of well-known and characterized *chitwin* representatives (Table 1.1). Every compound of Table 1.1 has two asymmetric carbons with the same chirality [22-25]. On the other hand, all of them have their *meso* (or *internally heterochiral*) counterpart. Of the ten isomers of linear hexitols, four are *chitwin* (D-and L-mannitol, D- and L-iditol) (Table 1.2) two are *meso* (allitol and galactitol) and four are based on an *irregular distribution* of their chiral centers {D-glucitol (L-gulitol) is (2S,3R,4R,5R)-hexahydroxy-hexane}. *Idem* for the ten linear isomers of aldaric acids (Table 1.2). Many other polyols and aldaric acids, especially those with an even number of C atoms and hydroxyl functions, present a similar molecular variety of their isomers. Also, there was a tentative to extend this concept to C<sub>8</sub> and C<sub>9</sub> [26-28].

Table 1.1: Chitwin molecules as represented by C<sub>4</sub> group.



		]	R			Chitwin compound
<b>R</b> 1	R2	R3	R4	R5	R6	
-COOH	-H	-OH	-OH	-H	-COOH	(2R,3R)-Tartaric acid (chitwin 1.1)
-COOH	-OH	-H	-H	-OH	-COOH	(2S,3S)-Tartaric acid (chitwin 1.2)
-H <sub>2</sub> C-OH	-H	-OH	-OH	-H	-H <sub>2</sub> C-OH	(2S,3S)-Threitol (chitwin 1.3)
-H <sub>2</sub> C-OH	-OH	-H	-H	-OH	-H <sub>2</sub> C-OH	(2R,3R)-Threitol (chitwin 1.4)
-CH <sub>3</sub>	-H	-OH	-OH	-H	-CH <sub>3</sub>	(2S,3S)-2,3-Butanediol (chitwin 1.5)
-CH <sub>3</sub>	-OH	-H	-H	-OH	-CH <sub>3</sub>	(2R,3R)-2,3-Butanediol (chitwin 1.6)
$-H_2C-C_6H_5$	-OH	-H	-H	-OH	$-H_2C-C_6H_5$	(2R,3R)-1,4-Diphenyl-2,3-butanediol
						(chitwin 1.7)
$-H_2C-C_6H_5$	-H	-OH	-OH	-H	$-H_2C-C_6H_5$	(2S,3S)-1,4-Diphenyl-2,3-butanediol
						(chitwin 1.8)
$-C(C_6H_5)_2-OCH_3$	-H	-OH	-OH	-H	$-C(C_6H_5)_2-OCH_3$	(2R,3R)-1,1,4,4,-Tetraphenyl-1,4-
						dimethoxy-2,3-butanediol (chitwin 1.9)

**1.2. 5-Ketohexoses** of some ketohexoses (fructose, sorbose) and their  $1,\omega$ -diphosphates (5-ketofructose-1,6-diphosphate) are *chitwin* molecules [29-32].

## 1.3. Dimeric saccharides

Fischer prepared other *chitwin* compounds by a doubling reaction of glycosyl bromides: trehalose, isotrehalose,  $\beta$ , $\beta$ -dicellobiose,  $\beta$ , $\beta$ -dilactose [33-36]. Hence, all homogenous unreducing disaccharides having the same ring and



glycosidic configuration are *chitwin* representatives. Trehalose ( $\alpha$ -D-glucopyranosyl-1,1'- $\alpha$ -D-glucopyranose) was found *per se* and as a 6,6'-diester of mycolic acid (cord factor) [37] (Fig. 1.1). Isotrehalose ( $\beta$ -D-glucopyranosyl-1,1'- $\beta$ -D-glucopyranose), with the two sugar residues linked by a spacer, was found in carotenoid glycosides (see polyprenyl lipids), lignans and cyclobutane alkaloids (see below). An isomer of *chitwin* tetrasaccharides synthesized by Fischer, based on gentiobiose, was found in crocin [38]. On the other hand, all the ether dimers of reducing saccharides linked to the same carbon, belong to this subgroup (Fig. 1.1). Till now, some representatives of 2,2'dimer ether of  $\beta$ -D-glucopyranose, linked directly, in cinnacasolide C [39], or of 6,6'-dimer ether, linked by a spacer in monochaetin [40], sagerinic acid [41] and stachysetin [42] (see below).

Table 1.2: *Chitwin* molecules as represented by C<sub>6</sub> group.



						-				
R									Chitwin compound	
<b>R</b> 1	R2	R3	<b>R4</b>	R5	<b>R6</b>	<b>R7</b>	<b>R8</b>	R9	R10	_
-H <sub>2</sub> C-OH	-OH	-H	-OH	-H	-H	-OH	-H	-OH	-H <sub>2</sub> C-OH	(2R,3R,4R,5R)-Mannitol (chitwin 1.10)
-H <sub>2</sub> C-OH	-H	-OH	-H	-OH	-OH	-H	-OH	-H	-H <sub>2</sub> C-OH	(2S,3S,4S,5S)-Mannitol (chitwin 1.11)
-H <sub>2</sub> C-OH	-OH	-H	-H	-OH	-OH	-H	-H	-OH	-H <sub>2</sub> C-OH	(2R,3S,4S,5R)-Iditol (chitwin 1.12)
-H <sub>2</sub> C-OH	-H	-OH	-OH	-H	-H	-OH	-OH	-H	-H <sub>2</sub> C-OH	(2S,3R,4R,5S)-Iditol (chitwin 1.13)
-COOH	-OH	-H	-OH	-H	-H	-OH	-H	-OH	-COOH	(2S,3S,4S,5S)-Mannaric acid (chitwin 1.14)
-COOH	-H	-OH	-H	-OH	-OH	-H	-OH	-H	-COOH	(2R,3R,4R,5R)-Mannaric acid (chitwin 1.15)
-COOH	-OH	-H	-H	-OH	-OH	-H	-H	-OH	-COOH	(2S,3R,4R,5S)-Idaric acid (chitwin 1.16)
-COOH	-H	-OH	-OH	-H	-H	-OH	-OH	-H	-COOH	(2R,3S,4S,5R)-Idaric acid



Figure 1.1: Unreducing oligosaccharides and reducing dimer ethers of saccharides as chitwin representatives

## 2. Amino Acids

It was Vickery [43] who noticed *chitwin* molecules as a distinct subgroup, and got involved systematically in these compounds, including their chemical nomenclature [44]. He defined them as "symmetrical substances which have identical asymmetric structures at both ends of a chain". Vickery showed that other classes of compounds, especially amino acids, may also submit to this principle.

Amino acids per se or as derivatives present three forms, all including representatives of chitwin molecules:

## 2.1. Diamino-dicarboxylic acids

Cystine and its higher homologues,  $\alpha_{,\epsilon}$ -diaminopimelic acids and other similar linear diamino-dicarboxylic acids, and lanthionine (Fig. 2.1).





**2.2.** Diketopiperazines and derivatives formed of aminoacids with the same configuration. Of the 20 common aminoacids, 19 produce *chitwin* diketopiperazines and derivatives (Tab. 2.1).

Table 2.1: 2,5-Diketopiperazines of natural aminoacids, as chitwin molecules

	$\begin{array}{c} H \\ HN \\ HN \\ HN \\ 0 \\ R \\ H \\ \end{array}$					
	pyperazines (c	hitwin 2.4)				
R	2,5-Diketopiperazine	R	2,5-Diketopiperazine			
-CH <sub>3</sub>	Ala (chitwin 2.5)	-CH <sub>2</sub> -CH <sub>2</sub> -CONH <sub>2</sub>	Gln (chitwin 2.14)			
-CH(CH <sub>3</sub> ) <sub>2</sub>	Val (chitwin 2.6)	-CH <sub>2</sub> -COOH	Asp (chitwin 2.15)			
-CH <sub>2</sub> -CH(CH <sub>3</sub> ) <sub>2</sub>	Leu (chitwin 2.7)	-CH <sub>2</sub> -CONH <sub>2</sub>	Asn (chitwin 2.16)			
-CH(CH <sub>3</sub> )-CH <sub>2</sub> -CH <sub>3</sub> (3S)	Ile (chitwin 2.8)	-CH <sub>2</sub> -(4-OH)Phenyl	Tyr (chitwin 2.17)			
$-CH(OH)-CH_3(3S)$	Thr (chitwin 2.9)	-CH <sub>2</sub> -Phenyl	Phe (chitwin 2.18)			
-CH <sub>2</sub> -OH	Ser (chitwin 2.10)	-CH <sub>2</sub> -Imidazolyl	His (chitwin 2.19)			
-CH <sub>2</sub> -SH	Cys (chitwin 2.11)	-CH <sub>2</sub> -Indolyl	Trp (chitwin 2.20)			
-CH <sub>2</sub> -CH <sub>2</sub> -S-CH <sub>3</sub>	Met (chitwin 2.12)	-(CH <sub>2</sub> ) <sub>2</sub> -guanidyl	Arg (chitwin 2.21)			
-CH <sub>2</sub> -CH <sub>2</sub> -COOH	Glu (chitwin 2.13)	$-(CH_2)_3-NH_2$	Lys (chitwin 2.22)			

**2.3. Homogenous diesters of amino acids**, their prototype being amino acid peroxydes and anhydrides. (For lysine diester with a spacer see carotenoid esters).

Cystine and diaminopimelic acid have been introduced in this group by Vickery [43] and lanthionine was introduced by us on the same biochemical basis. Cystine (Cys-Cys) may suffer a reducing reaction, cysteine (Cys) being the



reduced form. All three amino acids – cysteine, diaminopimelic acid, lanthionine, fully exemplify the type of indirect biosynthesis, every dimerization reaction being omitted [45], except conversion of Cys to Cys-Cys. This reaction is characteristic to all oligo- and polypeptides containing Cys (see 3. Coenzymes and 4. Homomeric Proteins).

2,5-Diketopiperazines were discovered by E. Fischer [46]. All possible forms of homogenous (LL, DD) and mixed (D and L) as well as of different amino acids, were synthesized and discovered in natural materials [47-52]. 2,5-Diketopiperazines formed of different amino acids are important since their doubling as chiral molecules leads to *chitwin* ones (see 6. Heterocyclic derivatives of 2,5-diketopiperazines). Cyclo(L-Val-L-Val) and cyclo(L-Val-D-Val) were synthesized in view of their comparative oxidation with dioxiranes [53]. Cyclodipeptide synthases were discovered as a novel enzyme family that employs aminoacyl-tRNAs as substrates for 2,5-diketopiperazine synthesis. A cyclodipeptide synthase of *Streptomyces noursei*, AlbC, uses aminoacyl-tRNAs as substrates to catalyze the formation of cyclo-L-Phe-L-Leu [54]. A number of 51 cyclodipeptide synthases were analyzed concerning their substrate specificity, and the conclusion was that they use 17 proteinogenic amino acids [55]. Two such enzymes of *Nocardiopsis* sp., NozA and NcdA, catalyze cyclo-L-Trp-L-Trp biosynthesis from tryptophanyl-tRNA, being outstandingly specific [56].

#### 3. Coenzymes

Coenzymes included in the subgroup of *chitwin* molecules are coenzyme A, 4-phosphopantetheine and glutathione, all in oxidized state (Fig. 3.1) [45]. Multiple biochemical and physiological reasons determined vast and detailed research about all three compounds. Coenzyme A is involved in the metabolism of organic acids and can serve also as a hydrogen carrier (Fig. 3.1). Phosphopantetheine, a fragment of coenzyme A, similarly with the latter, possesses a thiol group in its molecule. Of this reason, it may function as a redox system and bind to cellular proteins. A well studied tripeptide undergoing this transformation is glutathione; in oxidized form glutathione is a chitwin molecule (Fig. 3.1). Glutathione was studied especially due to its implications in redox reactions, detoxification processes and glutathione cycle. The amino acid Cys suffers decarboxylation and becomes cysteamine. This amino thiol is a biochemical constituent of phosphopantetheine and coenzyme A.





#### 4. Homomeric Proteins

When amino acids are integrated in protein chains, their amino and carboxyl groups of C- $\alpha$  constitute the peptide backbone, and manifest themselves in a totally different manner, in comparison with free state. Physical-chemical character of amino acids in this new state is predominantly expressed by their side chains (tails). They allow a variety of linkages, some of them relatively weak but significant in number (Van der Waals, hydrogen bonds), others strong, and of this reason easily approached: covalent (disulfuric), ionic, etc. Disulfuric bonds are due exclusively to Cys, a relatively widespread natural constituent of proteins. Hence this amino acid is an important agent for the association of proteins in oligomeric constructions, by disulfide bridges. Being formed of amino acids that are chiral, proteins are also chiral. And the association of two (or more) chiral structures of the same handedness can produce only a *chitwin* system and not a symmetrical one [57-60]. Consequently, homooligomeric structures or homomeric complexes are *chitwin*, rather than symmetric. The homomeric complexes are *chitwin* rather than symmetric of two reasons: (1) the constituent amino acids of proteins have L-configuration; (2) the sense of monomeric protein helices is of the same handedness. And the answer to a question raised by Metzler and Metzler [45] is this one: two chromatids coiled with opposite handedness form a *symmetric* system and snail shells or flowers [18,19] with both right and left handedness are also *symmetric*.

### 5. Lipids

Lipids are represented as *chitwin* molecules (Fig. 5.1) by cardiolipin [61], bis-phosphatidyl-dihydroxy acetone and other similar compounds. However, a doubling reaction of every lipidic chiral molecule would produce a *chitwin* structure. It was shown that when cholesterol, a chiral molecule with eight asymmetric C atoms, is put in the presence of concentrated sulfuric acid, a dimer, *bicholestadiene, is formed, in which cholesterol kept seven* asymmetric C atoms [62]. Bicholestadiene is a veritable chitwin compound. On the other hand, enantiomeric cholesterol has been synthesized [63]. It might be supposed that if racemic cholesterol is mixed with sulfuric acid, three types of bicholestadiene are formed, two chitwin and one symmetric.



Figure 5.1: Chitwin molecules represented by lipids

#### 6. Heterocyclic Derivatives of 2,5-Diketopiperazines

Compounds of this group manifest a variety of biological activities. The relatively loose diketopiperazine is closed to a tighter structure, both *chitwin* molecules [52]. Moreover, their lipophilic character is increased by methylation or prenylation (Fig. 6.1). Their meso isomers follow a similar metabolic path. Some of them have antibiotic activity, i. e., they are interspecies mediators. They are especially derivatives of substituted diketopiperazines, and are implicitly biosynthesized of amino acids. Only those that belong to *chitwin* molecules will be discussed here.

Diketopiperazines of two different amino acids are just chiral. Their doubling make them *chitwin*. The following *chitwin* dimers of mixed 2,5-diketopiperazines are known: cyclo-L-Phe-L-Trp: ditryptophenaline, WIN 64821 and *ent*-WIN 64821; cyclo-L-Trp-L-Ala and -S-S- bridges: (+)-11,11'-Dideoxyverticillin A; cyclo-L-Trp-L-Ser and -S-S- bridges: epidithiodiketopiperazine-1, epidithiodiketopiperazine-2; cyclo-L-Phe-L-Ser: vertihemiptellide A [52]. Producing of *chitwin* molecules by dimerization and a series of trimming reactions is very plausible for the following heterocyclic derivatives: cyclo-L-Phe-L-Phe and -S-S- bridges: scabrosin ester; cyclo-L-Glu-L-Glu: epicoccin C, epicoccin D; hematocin; mycoedyketopiperazine; epicoccin G; epicoccin H; alternarosin A [52].





Figure 6.1: Heterocyclic derivatives as chitwin molecules

#### 7. Polyprenyl (Isoprenoid) Compounds

Polyprenyl (isoprenoid) compounds present two facets as representatives of *chitwin* molecules:

**7.1. Perhydro optically active hydrocarbons**: squalane, perhydrolycopene, perhydrocrocetin, perhydronorbixin, perhydrocarotene, etc., [64].

7.2. Isoprenoid compounds per se or glycosylated or esterified with amino acids (Fig. 7.1).



Figure 7.1: Carotenoids and their derivatives as chitwin molecules



Carotenoids play the role of photons modulators in biochemistry and physiology. This biochemical role is shared with relatively few other molecules. Carotenoids are biosynthesized by a dimerization (condensation) reaction of digeranyl diphosphate [65-67]. The unsaturation of produced hydrocarbons (phytoene,  $\beta$ -carotene, lycopene) is stepwise brought to a conjugated system and then submitted to a variety of chemical transformations – hydroxylation, chain lenghthening or oxidative cleavage, epoxidation, glycosylation, amino acylation, etc., [68-70]. Carbohydrates currently used for carotenoids glycosylation are D-glucose [71] and L-rhamnose [72]. Many other *chitwin* carotenoids are known: crocin bis-glycosylated with  $\beta$ -(2- $\beta$ -cellobiosyl)gentiobiose [73],  $\gamma$ , $\gamma$ -carotene [74], astaxanthin [75],  $\beta$ , $\beta$ -carotene-2,2'-diol [76], bacterioruberin and bacterioruberin-diglucoside [77], auroxanthin ((8R,8'R)-, (8S,8'S)-) [78], bisanhydrobacyerioruberin [79]. More than 600 different carotenoid structures are known [80] and the question how many of these are *chitwin* will be answered quite soon.

#### 8. Lignans and Neolignans



Figure 8.1: Lignans and neolignans as kitwin molecules

The word lignan was coined by Haworth [81] as an unequivocal term concerning the vegetable origin of these compounds. Lignans and neolignans are typically dimerization products of monolignols, with a wide distribution in higher plants. Dimerization of monolignols is a remarkably versatile reaction: both head to head and head to tail linkages lead to lignans, the atoms involved being in both cases C-8 and C-8'. When linkages other than 8-8' are formed, e. g., 3-3' (or 5-5'), 8-O-4', and 8-3' (8-5'), neolignans are produced (Fig. 8.1). Most of the lignans and neolignans are optically active, some of them are even *chitwin* and others are *meso*. Metabolic interchangeability of



the three categories – *meso*, *chitwin* and based on *irregular distribution of chiral centers* – is well illustrated by lignans and neolignans. Structural variety of lignans and neolignans is increased yet by the diversity of structures forming aromatic rings and side chains. Of ten subtypes of classical lignans, six can provide chitwin molecules: dibenzocyclooctadiene, dibenzylbutane, dibenzylbutyrolactol (with the two alcoholic groups blocked by glycosylation, esterification or methylation), furofuran, 2,5-diaryltetrahydrofuran, 3,4-dibenzyl-tetrahydrofuran [82]. Diarylcyclobutane, of neolignan subtypes,provide a diversity of *chitwin* and *meso* derivatives (Fig. 8.1).

The number of biochemical precursors of metabolites produced by dimerization is relatively low in comparison with the number of final products:

- The major monolignols involved in the production of lignans are: coniferyl alcohol, (*E*)-isoeugenol, (*E*)-5-methoxyisoeugenol, and sinapyl alcohol [83-85]. Anol was converted, in the presence of (+)-larreatricin hydroxylase, a polyphenol oxidase from the creosote bush (*Larrea tridentata*), to the following metabolites: larreatricin, 3-hydroxy-larreatricin, 3,3'-dihydroxy-larreatricin, 3,3'-didemethoxyverrucosin, 3'-hydroxy-larreatricin, nordihydroguaiaretic acid, 3,3'-didemethoxy nectandrin B, 8'-*epi*-larreatricin, 3'-hydroxy-8'-*epi*-larreatricin; 3-O-Me-nor-dihydroguaiaretic acid, dihydroguaiaretic acid [86].
- Head to tail dimerization of coniferyl alcohol produces either (+)- or (-)-pinoresinol, the two enantiomers being biosynthesized in a characteristic ratio by different plants organs. The two enantiomers are both *chitwin* molecules and they are either glucosylated or converted to other metabolites, enterodiols being terminal points. Bis-β-D-Glcp glycosides of (+)- and (-)-pinoresinol and of (+)- and (-)-seoisolariciresinols are *chitwin*. The same type of dimerization of sinapyl alcohol leads to syringaresinols and yangambin [39].
- 3. Dimerization of cinnamic acid presents two alternatives [87]: head to tail and head to head. All isomers produced by head to tail dimerization are achiral: α-truxillic acid due an alternative axis of symmetry, and the others γ-, epi-, ε-, peri-, due to some planes of symmetry. All are constituent of alkaloids. Doubling of chiral molecules on these acids produces esters that are neither *chitwin* nor meso: incarvillateine [88], nitroxideincarvillateine, α-truxilline [89], santiaguine [90], thesine [91], hoveine, γ-truxilline, epi-truxilline, ε-truxilline, peri-truxilline. Head to head dimerization of cinnamic acid gives three types of acids: *chitwin* (δ- and μ-truxinnic acids), *meso* (β-truxinic and ω-truxillic acids), acids with an *irregular distribution of chiral carbons* (neo- and ζ-truxinic acids). Doubling of chiral compounds on *chitwin* acids produces *chitwin* molecules: nigramide R, sceptrin, stachysetin [42], sagerinic acid [41], monochaetin [40], δ-truxilline, μ-truxilline, ω-truxilline, 4,4'-dimethoxy-β-truxinic acid so overturns their status without making them chitwin: β-truxilline, α-truxilline, 4,4'-dimethoxy-β-truxinic acid catalpol diester. Doubling of chiral molecules on acids that are neither chitwin nor meso, keeps their status: neo-truxilline, ζ-truxilline [92].

## 9. Palindromes

Scholars approaching palindromes in a cultural perspective will be surprised to find them in two completely different, extreme positions: on one hand artistic (literary), and the scientific one, on the other hand. In fact they offered artistic delight before delivering scientific satisfaction, and over two millennia elapsed between the two facets.

A palindrome is a word, verse or sentence that is the same when read forward or backward, letter by letter. The inventor (discoverer) of palindromic writing was Sotades of Maroneea (in Thrace) [93,94], a Greek poet and satirist of the third century B. C. Being read in two directions, palindromes cause a double effect and Sotades had to pay with his life for a palindromic satire directed against the despot of the time [95].

Palindromes as biochemical phenomena were discovered in the second half of the twentieth century. Palindromic sequences occur frequently in DNA and have been often associated with functionally important locations such as replication points or operator sites [45]. They were detected soon after the discovery of DNA structure, due to some cruciform appearances containing intrastrand base pairing of the self-complementary sequence [96,97]. Then the DNA strands were sequenced [97,98] and in this way palindromic messages became quite explicit as *chitwin* structures (Fig. 9.1).



# A-C-G-T-A-A-G-C-G-C-T-T-A-C-G-T

# T-G-C-A-T-T-C-G-C-G-A-A-T-G-C-A

Figure 9.1: Palindromic sequence as chitwin structure

### 10. Is There a Chitwin (Bio)chemical Force?

It's a general principle that symmetrical joining of two chiral molecules leads to a *chitwin* chemical system. Steroid dimers have been isolated from living matter. As a result, steroid dimers – *chitwin* molecules – linked by a flexible diyne spacer have been synthesized (Fig. 10.1) [99].



Cholesterol diyne dimer (chitwin 10.1)

### Figure 10.1: Cholesterol diyne dimer as a chitwin structure

More independent research groups have undoubtedly established that cholesterol, when mixed with lipids that are common constituents of cellular membrane – phospholipids [100], dipalmitoylphosphatidylcholine [101], phosphatidylcholine or phosphatidylethanolamine [102], – forms dimers, i. e., *chitwin* structures. Moreover, attaching of a series of residues to hydroxyl group [103] or a fluorescent group on lateral chain [104,105] do not affect this behaviour. On the other hand, cholesterol induces dimerization of  $\beta$ 2-adrenergic receptor, the result being a *chitwin* construction [106]. Saposin B, the nonenzymatic glycosphingolipid activator protein, is destined to facilitate the hydrolysis of cerebroside sulfates (sulfatides) within the lysosome [107]. Human saposin B behaves as a shell-like dimer consisting of a monolayer of  $\alpha$ -helices enclosing a large hydrophobic cavity. The helices of saposin B are repacked into a tertiary arrangement to form the homodimer, and the latter is a *chitwin* structure.

## 11. Conclusions

*Chitwin* molecules are chiral structures formed of two identical halves, i. e., two sets of asymmetric C of the same chirality and two identical sets of chemical functions. They are homochiral with one another and are also internally homochiral. *Chitwin* structures have a unique, exceptional quality: no matter which of their half suffer chemical transformations, the result is the same. Doubling of a chiral molecule by its symmetrical joining, either directly by itself (trehalose, isotrehalose, 2,2'-di- $\beta$ -D-glucopyranosyl ether, cystine, 2,5-diketpiperazines of amino acids, glutathione and coenzyme A, both in oxidized state, lignans) or by an achiral (crocin, di- $\beta$ -D-glucopyranosyl-dihydroxy lycopene) or chiral spacer (diglucosides of zeaxanthin, decaprenoxanthin, astaxanthin, pinoresinol, secoisolariciresinol, stachysetin,  $\mu$ -truxilline) leads to a *chitwin* system. When this symmetrical doubling is made on a *meso* frame ( $\alpha$ -truxillic acid, meso-lignans), *meso* status is abolished, without becoming *chitwin*; a kind of hybrid product is rather obtained.

*Chitwin* molecules present a remarkable structural variety, they are well represented among the main classes of natural compounds and they are interchangeable with achiral, meso and chiral compounds. Due to these qualities, they constitute a real, essential, unique and trustworthy network in living matter.

### References

1. Pasteur, L. (1848). Memoire sur la relation qui peut exister entre la forme cristalline et la composition chimique, et sur la cause de la polarisation rotatoire. Comptes rendus des seances de l'academie des sciences, 26:535-538.



- 2. Pasteur, L. (1848). Mémoire sur la relation qui peut exister entre la forme crystalline et la composition chimique, et sur la cause dela polarization rotatoire. Comptes rendus des seances de l'academie des sciences, 26:33-38.
- 3. van 't Hoff, J. H. (1874). A suggestion looking to the extension into space of the structural formulas at present used in chemistry. And a note upon the relation between the optical activity the chemical constitution of organic compounds. Archives neerlandaises des sciences exactes et naturelles, 9:445-454.
- 4. Le Bel, J. A. (1874). Sur les relations qui existent entre les formules atomiques des corps organiques et le pouvoir rotatoire de leurs dissolutions. Bulletin de la societe chimique de France, 22:337-347.
- 5. Hoffmann, R., & Laszlo, P. (1991). Representation in Chemistry. Angewandte Chemie, 30:1-16.
- 6. Fischer, E. (1891). Ueber d. und i. Mannozuckersäure. Berichte der deutschen chemischen Gesellschaft, 24:539-546.
- 7. Fischer, E. (1891). Ueber die Configuration des Traubenzuckers und seiner Isomeren. Berichte der deutschen chemischen Gesellschaft, 24:1836-1845.
- 8. Fischer, E. (1891). Ueber die Configuration des Traubenzuckers und seiner Isomeren. II. Berichte der deutschen chemischen Gesellschaft, 24:2683-2687.
- 9. Fischer, E. (1896). Configuration der Weinsäure. Berichte der deutschen chemischen Gesellschaft, 29:1377-1383.
- 10. Bijvoet, J. M., Peerdemann, A. F., & van Bommel, A. J. (1951). Determination of the absolute configuration of optically active compounds by means of X-rays. Nature, 168:271-272.
- Tilden, W. A. (1921). Famous Chemists, The Men and their Work, George Routledge & Sons, London, pp. 52-62.
- 12. Wisniak, J. (2009). Carl Wilhelm Scheele. Revista CENIC Ciencias Químicas, 40(3):165-173.
- 13. Fischer, H. O. L., & MacDonald, D. L. (1951). Carbohydrate chemistry. Annual Revue of Biochemistry, 20:43-66.
- 14. Iga, D. P. (2018). Basic Principles of the Strategy Concerning the Elucidation of Configuration of Chiral Centers of Linear Isomeric Aldohexoses. Foundations of Chemistry, 20(1):31-41.
- 15. Kelvin, W. T. (1904). Baltimore Lectures on Molecular Dynamics and the Wave Theory of Light, C. J. Clay, London, pp. 602-642.
- Prelog, V. (2006). Chirality in Chemistry. Nobel Lecture, December 12, 1975. Croatica Chemica Acta, 79(3):XLIX-LVII. © The Nobel Foundation 1975.
- 17. Cronin, J., & Reisse, J. (2005). 3. Chirality and the Origin of Homochirality. In Lectures in Astrobiology, (Gargaud, M., Barbier, B., Martin, H., Reisse, J., eds.) Springer-Verlag, London, Vol. 1, pp. 73-114.
- 18. Jesson, L. K., & Barrett, S. C. H. (2002). Solving the puzzle of mirror-image flowers. Nature, 417:707.
- Jesson, L. K., & Barrett, S. C. H. (2003). The Comparative Biology Of Mirror-Image Flowers. International Journal of Plant Sciences, 164 (5 Suppl.):S237-S249.
- 20. Marazzi, B., & Endress, P. K. (2008). Patterns and Development of Floral Asymmetry in Senna (Leguminosae, Cassiinae). American Journal of Botany, 95(1):22-40.
- Schilthuizen, M., Craze, P. G., Cabanban, A. S., Davison, A., Stone, J., Gittenberger, E., & Scott, B. J. (2007). Sexual selection maintains whole-body chiral dimorphism in snails. Journal of Evolutionary Biology, 20:1941-1949.
- 22. Rubin, L. J., Lardy, H. A., & Fischer, H. O. L. (1952). Synthesis of the Optically Active Enantiomorphic 2,3-Butanediols. Journal of the American Chemical Society, 74(2):425-428.
- 23. Hill, R. K., & Bradberry, T. F. (1982). Absolute configuration of (+)-1,4-diphenyl-2,3-butanediol. Experientia, 38(1):70-71.
- 24. De Mas, C., Jansen, N. B., & Tsao, G. T. (1988). Production of optically active 2,3-butanediol by Bacillus polymyxa. Biotechnology and Bioengineering, 31(4):366-377.



- 25. Nakayama, K., & Rainier, J. D. (1990). Synthesis of (2R,3R)-1,4-dimethoxy-1,1,4,4-tetraphenyl-2,3butanediol: a new C<sub>2</sub>-symmetric vicinal diol from dimethyl L-tartrate. Tetrahedron, 46(12):4165-4170.
- 26. Fischer, E., & Passmore, F. (1890). Ueber kohlenstoffreichere Zuckerarten aus d. Mannose. Berichte der deutschen chemischen Gesellschaft, 23:2226-2239.
- Hann, R. M., Maclay, W. D., Knauf, A. E., & Hudson, C. S. (1939). Relations between Rotatory Power and Structure in the Sugar Group. XXXI. The Configuration of D-α,α-Mannooctose (D-Manno-L-mannooctose). Journal of the American Chemical Society, 61(5):1268-1269.
- 28. Hudson, C. S. (1941). Emil Fischer's Discovery of the Configuration of Glucose. Journal of Chemical Education, 18:353-357.
- 29. Terada, O., Suzuki, S., & Kinoshita, S. (1961). Occurrence of 5-Ketofructose during Kojic Acid Formation from Sorbose by Acetobacter Species. Agricultural and Biological Chemistry, 25(11): 871-872,.
- 30. Yamada, Y., Iizuka, K., Aida, K., & Uemura, T. (1966). L-Sorbose Oxidase from Trametes sanguinea. Agricultural and Biological Chemistry, 30(1):97-98.
- Healy, M. J., & Christen, P. (1972). Reaction of the carbanionic aldolase-substrate intermediate with tetranitromethane. Identification of the products, hydroxypyruvaldehyde phosphate and D-5-ketofructose 1,6-diphosphate. Journal of the American Chemical Society, 94(22):7911-7916.
- 32. Ameyama, M., Chiyonobu, T., & Adachi, O. (1974). Purification and Properties of 5-Ketogluconate Reductase from Gluconobacter liquefaciens. Agricultural and Biological Chemistry, 38(7):1377-1382.
- 33. Fischer, E., & Armstrong, E. F. (1902). Ueber die isomeren Acetohalogen-Derivate der Zucker und die Synthese der Glucoside. Berichte der deutschen chemischen Gesellschaft, 35(1):833-843.
- 34. Fischer, E., & Armstrong, E. F. (1902). Synthese einiger neuer Disaccharide. Berichte der deutschen chemischen Gesellschaft, 35(3):3144-3153.
- 35. Fischer, E., & Delbrück, K. (1909). Synthese neuer Disaccharide vom Typus der Trehalose. Berichte der deutschen chemischen Gesellschaft, 42(2):2776-2785.
- 36. Armstrong, E. F. (1924). The Carbohydrates and The Glucosides, Longmans, London, pp. 151-154.
- 37. Noll, H., Bloch, H., Asselineau, J., & Lederer, E. (1956). The chemical structure of the cord factor of Mycobacterium tuberculosis. Biochimica et Biophysica Acta, 20(2):299-309.
- 38. Britton, G., Liaaen-Jensen, S., & Pfander, H. (2004). Carotenoids, Springer, Basel AG.
- 39. Zhao, J., & Ma, J.-S. (2016). Phytochemicals and Biological Activities of the Genus Cinnamomum. Journal of Pharmacognosy and Phytochemistry, 4(1):27-34.
- 40. Isaza, J. H., Ito, H., & Yoshida, T. (1999). Monochaetin, a Di-hyperin Ester of Tetrahydroxy-μ-truxinic Acid from Monochaetum multiflorum. Chemical and Pharmaceutical Bulletin, 47(10):1510-1511.
- 41. Lee, Y. H., Kim, B., Kim, S., Kim, M.-S., Kim, H., Hwang, S.-R., Kim, K., & Lee, J. H. (2017). Characterization of metabolite profiles from the leaves of green perilla (Perilla frutescens) by ultra high performance liquid chromatography coupled with electrospray ionization quadrupole time-of-flight mass spectrometry and screening for their antioxidant properties. Journal of Food and Drug Analysis, 25:776-788.
- 42. Zhang, F., & Jia, Y. (2009). Total Synthesis of (-)-Incarvilline and (-)-Incarvillateine. Tetrahedron, 65:6840-6843.
- 43. Vickery, H. B. (1957). Assignment of D L prefixes to the tartaric acids. Journal of Chemical Education, 34:339-341.
- Downey, P. F., & Black, S. (1957). A New Naturally Occurring Isomer of β-methyllanthionine. Journal of Biological Chemistry, 228:171-179.
- 45. Metzler, D. E., & Metzler, C. M. (2003). Biochemistry: the chemical reactions of living cells. Amsterdam: Elsevier.
- 46. Fischer, E. (1906). Synthese von Polypeptiden. XV. Berichte der deutschen chemischen Gesellschaft, 39(3):2893-2931.



- 47. Nitecki, D. E., Halpern, B., & Westley, J. W. (1968). A Simple Route to Sterically Pure Diketopiperazines. Journal of Organic Chemistry, 33(2):864-866.
- Kopple, K. D. & Ghazarian, H. G. (1968). A Convenient Synthesis of 2,5-Piperazinediones. Journal of Organic Chemistry, 33(2):862-864.
- Jung, M. E., & Rohloff, J. C. (1985). Organic Chemistry of L-Tyrosine. 1. General Synthesis of Chiral Piperazines from Amino Acids. Journal of Organic Chemistry, 50(24):4909-4913.
- 50. Cui, C.-B., Kakeya H., & Osada, H. (1996). Novel Mammalian Cell Cycle Inhibitors, Tryprostatins A, B and Other Diketopiperazines Produced by Aspergillus fumigatus. II. Physico-chemical Properties and Structures. Journal of Antibiotics, 49(6):534-540.
- 51. Huang, R., Zhou, X., Xuc, T., Yang, X., & Liu, Y. (2010). Diketopiperazines from Marine Organisms. Chemistry and Biodiversity, 7:2809-2829.
- 52. Borthwick, A. D. (2012). 2,5-Diketopiperazines: Synthesis, Reactions, Medicinal Chemistry, and Bioactive Natural Products. Chemistry Revues, 112:3641-3716.
- Annese, C., D'Accolti. L., Fusco, C., & Ciriaco F. (2016). Advances in Artificial Life, Evolutionary Computation and Systems Chemistry. WIVACE 2015. Communications in Computer and Information Science, vol 587. Rossi F., Mavelli F., Stano P., Caivano D. (eds) Springer, Cham.
- 54. Gondry, M., Sauguet, L., Belin, P., Thai, R., Amouroux, R., Tellier, C., Tuphile, K., Jacquet, M., Braud, S., Courçon, M., Masson, C., Dubois, S., Lautru, S., Lecoq, A., Hashimoto, S., Genet, R., & Pernodet, J. L. (2009). Cyclodipeptide synthases are a family of tRNA-dependent peptide bond-forming enzymes. Nature Chemical Biology, 5(6):414-420.
- Jacques, I. B., Moutiez, M., Witwinowski, J., Darbon, E., Martel, C., Seguin, J., Favry, E., Thai, R., Lecoq, A., Dubois, S., Pernodet, J.-L., Gondry M., & Belin, P. (2015). Analysis of 51 cyclodipeptide synthases reveals the basis for substrate specificity. Nature Chemical Biology, 11:721-727.
- James, E. D., Knuckley, B., Alqahtani, N., Porwal, S., Ban, J., Karty, J. A., Viswanathan, R., & Lane, A. L. (2016). Two Distinct Cyclodipeptide Synthases from a Marine Actinomycete Catalyze Biosynthesis of the Same Diketopiperazine Natural Product. ACS Synthetic Biology, 5(7):547-553.
- 57. Pereira-Leal, J. B., Levy, E. D., Kamp C., & Teichmann, S. A. (2007). Evolution of protein complexes by duplication of homomeric Interactions. Genome Biology, 8(4):R51.1-R51.12.
- Venkatakrishnan, A. J., Levy E. D., & Teichmann, S. A. (2010). Homomeric protein complexes: evolution and assembly. Biochemical Society Transactions, 38:879-882.
- 59. Kojic-Prodic, B., & Stefanic, Z. (2010). Symmetry versus Asymmetry in the Molecules of Life: Homomeric Protein Assemblies. Symmetry, 2:884-906.
- 60. Bergendahl, L. T., & Marsh, J. A. (2017). Functional determinants of protein assembly into homomeric complexes. Scientific Reports, 7: 4932.
- 61. Kim, J., Minkler, P. E., Salomon, R. G., Anderson, V. E., & Hoppel. C. L. (2011). Cardiolipin: characterization of distinct oxidized molecular species. Journal of Lipid Research, 52(1):125-135.
- 62. Rapoport, S. M., & Raderecht, H.-J. (1972). Physiologisch-chemisches Praktikum. Veb Verlag Volk Und Gesundheit, Berlin, pp. 290-291.
- 63. Crowder, C. M., Westover, E. J., Kumar, A. S., Ostlund, R. E., Jr., & Covey, D. F. (2001). Enantiospecificity of Cholesterol Function in Vivo. Journal of Biological Chemistry, 276:44369-44372.
- 64. Karrer, P., & Jucker, E. (1950). Carotenoids, Elsevier, Amsterdam.
- 65. Lagarde, D., & Vermaas, W. (1999). The zeaxanthin biosynthesis enzyme IS-carotene hydroxylase is involved in myxoxanthophyll synthesis in Synechocystis sp. PCC 6803. FEBS Letters, 454:247-251.
- Maresca, J. A., & Bryant, D. A. (2006). Two Genes Encoding New Carotenoid-Modifying Enzymes in the Green Sulfur Bacterium Chlorobium tepidum. Journal of Bacteriology, 188(17): 6217-6223.
- Li, F., Murillo, C., & Wurtzel, E. T. (2007). Maize Y9 Encodes a Product Essential for 15-cis-ζ-Carotene Isomerization. Plant Physiology, 144:1181-1189.



Chemistry Research Journal

- 68. Misawa, N., Satomi, Y., Kondo, K., Yokoyama, A., Kajiwara, S., Saito, T., Ohtani, T., & Miki, W. (1995). Structure and Functional Analysis of a Marine Bacterial Carotenoid Biosynthesis Gene Cluster and Astaxanthin Biosynthetic Pathway Proposed at the Gene Level. Journal of Bacteriology, 177(22): 6575-6584.
- Jackson, H. L., Cardounel, A. J., Zweier, J. L., & Lockwood, S. F. (2004). Synthesis, characterization, and direct aqueous superoxide anion scavenging of a highly water-dispersible astaxanthin-amino acid conjugate. Bioorganic & Medicinal Chemistry Letters, 14(15):3985-3991.
- 70. Ohmiya, A. (2011). Diversity of Carotenoid Composition in Flower Petals. Japan Agricultural Research Quarterly, 45(2):163-171.
- 71. Umeno, D., Tobias, A. V., & Arnold, F. H. (2005). Diversifying Carotenoid Biosynthetic Pathways by Directed Evolution. Microbiology and Molecular Biology Reviews, 69(1):51-78.
- 72. Nybraaten, G., & Liaaen-Jensen, S. (1974). Bacterial Carotenoids. XLIV. Zeaxanthin Mono- and Dirhamnoside. Acta Chemica Scandinavica B, 28:1219-1224.
- 73. Ahrazem, O., Rubio-Moraga, A., Jimeno, M. L., & Gómez-Gómez, L. (2015). Structural characterization of highly glucosylated crocins and regulation of their biosynthesis during flower development in Crocus. Frontiers in Plant Science, 6:971.
- 74. Andrewes, A. G., & Liaaen-Jensen, S. (1971). Fungal Carotenoids. VII. Synthesis of  $\beta$ , $\gamma$  and  $\gamma$ , $\gamma$ -carotene with terminal methylene groups. Acta Chemica Scandinavica B, 25(5):1922-1923.
- 75. Andrewes, A. G., Borch, G., Liaaen-Jensen, S., & Snatzke, G. (1974) Animal Carotenoids. 9. On the Absolute Configuration of Astaxanthin and Actinioerythrin. Acta Chemica Scandinavica B, 28:730-736.
- Buchecker, R., Eugster, C. H., Kjosen, H., Liaaen-Jensen, S. (1974). Algal Carotenoids. IX. Absolute Configuration of β,ε-Caroten-2-ol, β,β-Caroten-2-ol, and β,β-Carotene-2,2'-diol. Acta Chemica Scandinavica B, 28(4):449-452.
- Arpin, N., Liaaen-Jensen, S., & Trouilloud, M. (1972). Bacterial Carotenoids XXXVIII. C<sub>50</sub>-Carotenoids 9. Isolation of Decaprenoxanthin Mono- and Diglucoside from an Arthrobacter sp. Acta Chemica Scandinavica B, 26(6):2526-2528.
- 78. Asai, A., Terasaki, M., & Nagao, A. (2004). An Epoxide–Furanoid Rearrangement of Spinach Neoxanthin Occurs in the Gastrointestinal Tract of Mice and In Vitro: Formation and Cytostatic Activity of Neochrome Stereoisomers. Journal of Nutrition, 2237-2243.
- 79. Borch, G., Norgard, S., & Liaaen-Jensen, S. (1972). C50-Carotenpoids. 8. Circular dichroism and relative configuration of C50-carotenoids. Acta Chemica Scandinavica B, 26(1):402-403.
- 80. Brocks, J. J., & Schaeffer, P. (2008). Okenane, a biomarker for purple sulfur bacteria (Chromatiaceae), and other new carotenoid derivatives from the 1640 Ma Barney Creek Formation. Geochimica et Cosmochimica Acta, 72:1396-1414.
- 81. Haworth, R. D. (1937). Natural resins. Annual Report and Progress in Chemistry, 33:266-279.
- 82. Teponno, R. B., Kusari, S., & Spiteller, M. (2016). Recent advances in research on lignans and neolignans. Natural Product Reports, 33:1044-1092.
- Davin, L. B., & Lewis, N. G. (2000). Dirigent Proteins and Dirigent Sites Explain the Mystery of Specificity of Radical Precursor Coupling in Lignan and Lignin Biosynthesis. Plant Physiology, 123, 453-461.
- 84. Lopes, N. P., Yoshida, M., & Kato, M. (2004). Biosynthesis of tetrahydrofuran lignans in Virola surinamensis. Brazilian Journal of Pharmaceutical Sciences, 40(1):53-57.
- 85. Rahman, A., Katayama, T., Suzuki, T., & Nakagawa, T. (2007). Stereochemistry and biosynthesis of (+)lyoniresinol, a syringyl tetrahydronaphthalene lignan in Lyonia ovalifolia var. elliptica I: isolation and stereochemistry of syringyl lignans and predicted precursors to (+)-lyoniresinol from wood. Journal of Wood Science, 53:161–167.
- Cho, M.-H., Moinuddin, S. G. A., Helms, G. L., Hishiyama, S., Eichinger, D., Davin, L. B., & Lewis, N. G. (2003). (+)-Larreatricin hydroxylase, an enantio-specific polyphenol oxidase from the creosote bush



(Larrea tridentata). Proceedings of the National Academy of Sciences of the United States of America, 100(19):10641-10646.

- Atkinson, S. D. M., Almond, M. J., Hollins, P., Jenkins, S. L. (2003). The photodimerisation of the α- and β-forms of trans-cinnamic acid: a study of single crystals by vibrational microspectroscopy. Spectrochimica Acta Part A, 59:629-635.
- Chi, Y.-M., Nakamura, M., Zhao, X.-Y., Yoshizawa, T., Yan, W.-M., Hashimoto, F., Kinjo, J., & Nohara, T. (2005). A Monoterpene Alkaloid from Incarvillea sinensis. Chemical & Pharmaceutical Bulletin, 53(9):1178-1179.
- 89. De Simone, R., Margarucci, L., & De Feo, V. (2008). Tropane Alkaloids: An Overview. Pharmacologyonline, 1:70-89.
- 90. Dominguez, J., Mendez, M. R., & Ribas, I, (1956). Alkaloides de las Papilionaceas. XXIX. Estructura y sintesis de la santiaguina. Anales de la Real Sociedad Espanola de Fisica y Quimica, LII (2):133-137.
- 91. Arendaruk, A. P., Skoldinov, A. P., & Kharkevich, D. A. (1967). Studies in the cyclobutanedicarboxylic acid field iv. Synthesis of bis-quaternary salts of alkylamino esters of α-truxillic acid. UDC 615.785.3-015. Translated from Khimiko-Farmatsevticheskii Zhurnal, 4:3-8.
- 92. Mallette, J. R., & Casale, J. F. (2014). Rapid determination of the isomeric truxillines in illicit cocaine via capillary gas chromatography/flame ionization detection and their use and implication in the determination of cocaine origin and trafficking routes. Journal of Chromatography A, 1364:234-240.
- 93. Cameron, A. (1990). Two Mistresses of Ptolemy Philadelphus, GRBS, 31:287-311.
- 94. Tunny, J. A. (2000). Ptolemy "The Son" Reconsidered: Are There Too Many Ptolemies? from: Zeitschrift für Papyrologie und Epigraphik 131:83-92.
- 95. Mori, A. (2001). Personal Favor and Public Influence: Arete, Arsinoë II, and the Argonautica. Oral Tradition, 16(1):85-106.
- 96. Platt, J. R. (1955). Possible Separation of Intertwined Nucleic Acid Chains by Transfer-Twist. Proceedings of the National Academy of Sciences of the United States of America, 41:181-183.
- 97. Gellert, M., O'Dea, M. H., & Mizuuchi, K. (1983). Slow cruciform transitions in palindromic DNA. Proceedings of the National Academy of Sciences of the United States of America, 80:5545-5549.
- 98. Ellis, J. G., Llewellyn, D. J., Walker, J. C., Dennis E. S., & Peacock, W. J. (1987). The ocs element: a 16 base pair palindrome essential for activity of the octopine synthase enhancer. EMBO Journal, 6(11):3203-3208.
- Valdez-Garcia, R. M., Alarcon-Manjarrez, C., Arcos-Ramos, R., Flores-Alamo, M., & Iglesias-Arteaga, M. A. (2018). Synthesis and characterization of dimeric steroids based on 5-oxo-4,5-seco-yne units linked by a diyne spacer. Arkivoc, part iv:13-22.
- 100. Martin, R. B., & Yeagle, P. L. (1978). Models for Lipid Organization in CholesteroI-Phospholipid Bilayers Including Cholesterol Dimer Formation. Lipids, 13(9):594-597.
- 101. Harris, J. S., Epps, D. E., Davio, S. R., & Kezdy, F. J. (1995). Evidence for Transbilayer, Tail-to-Tail Cholesterol Dimers in Dipalmitoylglycerophosphocholine Liposomes. Biochemistry, 34:3851-3857.
- 102. Huang, J., & Feigenson, G. W. (1999). A Microscopic Interaction Model of Maximum Solubility of Cholesterol in Lipid Bilayers. Biophysical Journal, 76:2142–2157.
- 103. Yelamaggad, C. V., Shanker, G., Hiremath U. S., & Krishna Prasad, S. (2008). Cholesterol-based nonsymmetric liquid crystal dimers: an overview. Journal of Materials Chemistry C, 18:2927-2949.
- 104. Mukherjee, S., & Chattopadhyay, A. (1996). Membrane Organization at Low Cholesterol Concentrations: A Study Using 7-Nitrobenz-2-oxa-1,3-diazol-4-yl-Labeled Cholesterol. Biochemistry, 35:1311-1322.
- Mukherjee, S., & A. Chattopadhyay, (2005). Monitoring cholesterol organization in membranes at low concentrations utilizing the wavelength-selective fluorescence approach. Chemistry and Physics of Lipids, 134:79–84.



- 106. Prasanna, X., Chattopadhyay, A., & Sengupta, D. (2014). Cholesterol Modulates the Dimer Interface of the b2-Adrenergic Receptor via Cholesterol Occupancy Sites. Biophysical Journal, 106(6):1290-1300.
- 107. Ahn, V. E., Faull, K. F., Whitelegge, J. P., Fluharty, A. L., & Privé, G. G. (2003). Crystal structure of saposin B reveals a dimeric shell for lipid binding. Proceedings of the National Academy of Sciences of the United States of America, 100(1):38-43.

